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Predictive Significance of the Inflammatory Activities and GGT in Hepatocellular Carcinoma Development

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Received: 24/9/2025 Accepted: 13/10/2025 Abstract: This study evaluated whether gamma-glutamyltransferase (GGT) and several non-invasive inflammatory and fibrotic markers could predict the development of hepatocellular carcinoma (HCC). The research included 37 patients with HCC, 45 with liver cirrhosis, and 20 healthy individuals. Patients and Methods: Serum levels of AFP and GGT with the inflammatory markers (AAR, NLR, PLR and Pt) and fibrotic indices (APRI and FIB-4) were investigated in 82 adult HCC and liver cirrhotic patients in addition to 20 healthy individuals as non-disease control (NDC). Results: Our results showed that GGT, along with indices such as AAR, APRI, FIB-4, NLR, and PLR, were significantly elevated in HCC and cirrhotic patients compared to healthy controls. Alpha-fetoprotein (AFP) was the most accurate diagnostic marker (AUC = 0.972). Regression analysis indicated that ALT, INR, and platelet count could serve as early predictors of tumor development. Overall, the findings support the usefulness of combining inflammatory and fibrotic indices with traditional markers to improve early detection of HCC. Conclusion: AAR, NLR, FIB4, APRI and GGT as inflammatory and fibrotic indices play an important role in HCC and liver cirrhosis. ALT, INR and Pt count could be used as early predictors for HCC development.

keywords: AAR; APRI; AFP; FIB4; GGT

1.Introduction

Hepatocellular carcinoma (HCC) is a common malignancy in human [1]. HCC is one of the most common cancers worldwide and a major frequent cause of cancer mortality [2]. In Egypt, HCC is the 1st mortality related to cancer [3]. Only 10% to 20% of the HCCs can be surgically excised, although attended with a high frequency of recurrence [4].

HCC is a complex disease with multiple steps and associated with many risk factors and cofactors [5]. Most patients with HCC have a history of liver cirrhosis and chronic liver disease, with risk factors including infectious causes like hepatitis C virus and hepatitis B virus and noninfectious causes like nonalcoholic and alcoholic liver disease [3,6].

The incidence of HCC continues to rise despite the fast-paced period of direct acting

antiviral regimens (DAA) in the treatment of HCV [7]. Even when the virus has been successfully eradicated, individuals still experience chronic HCV complications such liver cirrhosis and HCC. Given the high rate of recurrence, postoperative HCC patients' long-term survival is inadequate. Therefore, there is a pressing need to uncover novel serological biomarkers with high accuracy and practicality for the early identification of HCC because serum α -Fetoprotein level detection of HCC is hampered by its low sensitivity [8].

The lack of readily available biomarkers that are both sensitive and specific is the main issue faced by physicians in HCC patient's management of HCC. Novel circulating indicators are highly needed to raise the rate of disease-free survival [9]. The most popular

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tumor marker for HCC patients identifying is serum alpha fetoprotein (AFP), which has also been shown to be able to predict prognosis [10]. Therefore, the aim of this study was to investigate the potential role of the inflammatory markers and fibrotic indices with GGT for predicting HCC earlier.

2. Materials and methods

Patients

The present study was conducted on 82 patients hospitalized for liver transplantation attending at the Gastro Intestinal Surgical Center (GISC) from August 2021 to April 2024 after approval of the Ethics Committee at the Faculty of Medicine, Mansoura University (Approval code MPD.21.04.66) and written informed consent was obtained from all participants. They were diagnosed according to the American Association for the Study of Liver Diseases (AASLD) guidelines, based on AFP > 400 and the presence of hepatic focal lesion(s) detected by liver ultrasound and confirmed by triphasic computed tomography and/or dynamic magnetic resonance imaging (MRI). The second group included patients with liver cirrhosis defined by clinical, biochemical, and imaging findings such as splenomegaly. They were followed up for 6 months to ensure the absence of HCC; 37 patients having HCC on top of liver cirrhosis (HCC-LC, 32 men and 5 women; median age 58yrs) and 45 liver cirrhotic patients (LC, 30 men and 15 women; median age 53yrs), in addition to 20 healthy individuals as control group (these individuals had no clinical history of hepatitis and no symptoms or signs of liver disease). Demographic data were collected from all participants.

Exclusion criteria included patients infected with HBV or HIV, patients with prior or metastatic HCC, patients with malignancies other than HCC, or patients aged less than 18 years old [11].

Sample collection and biochemical analysis

5 milliliters of venous blood were drawn from both patient's group and control, placed in plain tubes, blood was allowed to clot and the serum was separated by centrifugation at 4000 rpm for 15 minutes. Sera were immediately assessed for the biochemical parameters; ALT, AST, GGT, viral markers and the rest of serum

samples were stored at -80C° until used for analysis of AFP by ELISA.

Determination of noninvasive indices

Serum ALT and AST activities were calorimetrically determined according to the method described by Reitman and Frankel (1957), using a commercial kit supplied by Spin react, Santa Coloma, Spain.

The following ratios and indices (AAR, APRI, FIB4, PLR and NLR) were calculated as follow:

- **AAR** (AST to ALT Ratio) = AST/ALT. [12]
- **APRI** (AST to Platelet Ratio Index) was calculated using the following equation: (AST/upper limit of normal for AST) / platelet count ($\times 10^9$ /L) $\times 10$ [12].
- **FIB-4** = (Age in years \times AST) / (Platelet count $\times \sqrt{ALT}$), where AST and ALT are measured in U/L and platelets are in 10^9 /L. (a possible value of 0-10) [13].
- **NLR** (Neutrophil to Lymphocyte Ratio): Neutrophil / lymphocyte counts [14].
- **PLR** (Platelet to Lymphocyte Ratio): Platelet count / absolute lymphocyte count.

Statistical analysis

The collected data were analyzed and presented graphically using SPSS 20 (Statistical Package for Social Sciences). For continuous variables, descriptive statistics are reported as median and interquartile range (IQR). Kruskal-Wallis and Mann-Whitney U tests were employed to determine significant differences between non-parametric variables, with a p-value less than 0.05 considered statistically significant. Spearman's correlation coefficient was used for correlative analysis between variables. The area under receiver operating characteristic (ROC) curve (AUC) was utilized for assessment the diagnostic performance of biomarkers [15].

3. Results and Discussion

Result

The current case-control study included 102 participants were divided as follow, Group I consisting of 37 HCC-LC patients [32 males (86.5%) and 5 females (13.5%), with a median age of 58 years], Group II consisting of 45 LC

patients [30 males (66.7%) and 15 females (33.3%); with median age 53 years], and Group III consisting of 20 healthy subjects as Non-Disease control (NDC) group [18 males (90%) and 2 females (10%); with median age 34 years]. There was a significant difference in age ($\mathbf{p} < \mathbf{0.0001}$) and gender ($\mathbf{p} = \mathbf{0.033}$) as shown in **Table (1)**. Significant elevation of ALT was recorded in HCC patients compared to LC ($\mathbf{p} = \mathbf{0.007}$), between HCC and NDC ($\mathbf{p} < \mathbf{0.0001}$) and between LC and NDC ($\mathbf{p} = \mathbf{0.003}$). Higher expression was also detected in HCC and LC

compared to NDC as regards to AST and CRP (P< 0.05). Considered to AAR, INR, APRI and FIB-4, substantial increase was recorded in HCC and LC compared to NDC (p<0.0001). Enhanced level was also noticed in HCC and LC compared to NDC concerning PLR (p=0.002 and p=0.037), NLR (p=0.001 and p<0.0001) and GGT (p=0.002 and p=0.038), respectively). AFP showed higher activity (p<0.0001) in HCC and LC related to NDC and in HCC concerned to LC (Table 1).

Table (1) Clinicopathological and demographic characteristics of all study groups

	Group I(HCC-LC) N = 37	Group II (LC)N = 45	Group III (Non-Disease)N = 20	P Value	
M/F	32/5	30 /15	18/2	0.035	
Age (Yrs)	58 (54.5-59.5)	53 (43.5-59.0)	34 (25.0-44.75)	${}^{a}P=0.004$ ${}^{b,c}P<0.0001$	
AST (U/ml)	44 (31.0-55.5)	36(26.5-59.5)	21 (20.0-23.0)	$^{a}P = 0.152$ $^{b,c}P < 0.0001$	
ALT (U/ml)	28 (22.5-50.0)	23(20.0-30.0)	21 (20.0-21.0)	${}^{a}P=0.007$ ${}^{b}P<0.0001$ ${}^{c}P=0.003$	
Plt Count (10 ⁹ /L)	64.0 (46.0-106.0)	53.0(37.0-68.7)	218.9 (211.0-312.5)	${}^{a}P=0.038$ ${}^{b,c}P<0.0001$	
INR	1.2 (1.2-1.55)	1.4 (1.2-1.6)	1.0 (1.0-1.083)	${}^{a}P=0.078$ ${}^{b,c}P<0.0001$	
CRP (mg/L)	4.0 (4.0-11.8)	4.0 (3.0-11.5)	4.0(2.25-4.0)	${}^{a}P=0.598$ ${}^{b}P=0.001$ ${}^{c}P=0.009$	
APRI	1.77 (1.043-3.297)	1.805 (1.167-2.767)	0.245 (0.167-0.263)	$^{a}P = 0.625$ $^{b,c}P < 0.0001$	
FIB4	6.257 (4.289-12.337)	6.99 (4.58-10.25)	0.575 (0.319-0.665)	$^{a}P=0.678$ $^{b,c}P<0.0001$	
NLR	2.19 (1.279-3.64)	2.64 (1.377-3.595)	1.227 (0.908-1.46)	${}^{a}P=0.612$ ${}^{b}P=0.001$ ${}^{c}P<0.0001$	
AAR	1.385(1.109-1.763)	1.428 (1.1559-1.759)	1.05 (0.964-1.095)	$^{a}P=0.548$ $^{b,c}P<0.0001$	
PLR	67.27 (51.8-90.29) 80.95 (61.54-113.39) 88.77 (88.52-132.85)		${}^{a}P=0.233$ ${}^{b}P=0.002$ ${}^{c}P=0.037$		
AFP (ng/mL)	26.0 (4.65-68.5)	3.5.0 (2.395-5.05)	1.95 (0.93-2.0)	$^{a,b,c}P < 0.0001$	
GGT (U/L)	38.0 (26.0-68.0)	29.0 (22.0-66.0)	19.0 (14.0-36.0)	${}^{a}P=0.198$ ${}^{b}P=0.002$ ${}^{c}P=0.038$	
HCV infection +HCV Abs/-HCV RNA -HCV Abs/ -HCV RNA	34 /37 (91.9%) 3/37 (8.1%)	18/45 (40%) 27/45 (60%)	20/20 (100%)	P<0.0001	

a,b,c represent p<0.05 considered significant in HCC vs. LC, in HCC vs. NDC,in LC vs. NDC control, respectively.

Data are presented as median (Med) and interquartile range (IQR, 25^{th} - 75^{th})

Abbreviations: HCC-LC: hepatocellular carcinoma, LC: liver cirrhosis, NDC: non disease control, AST: aspartate

aminotransferase, *ALT*: Alanine aminotransferase, INR: international normalized Ratio, AAR: aspartate aminotransferase-to-alanine aminotransferase ratio, APRI: aminotransferase-to-platelet ratio index, FIB-4: Fibrosis-4 score, NLR: neutrophil lymphocyte ratio, PLR: platelet-tolymphocyte ratio, CRP: C-reactive protein, AFP: Alfa-fetoprotein.

Spearman correlation coefficient of GGT, AST and AFP (**Table 2, Fig 1**) displayed AFP was associated with AST, ALT, Pt and NLR; AST was associated with ALT, AAR, APRI, FIB-4, NLR and PLR. Also, GGT was significantly associated with ALT and, Pt. Regression analysis showed that AFP, ALT, Pt, and INR displayed substantial prediction for tumor development and may be employed as early markers of the onset of HCC (**Table 3**). The diagnostic performance of all biomarkers was studied by ROC curve analysis and only AFP demonstrated excellent accuracy (90.2%) with an AUC of 0.927 (**Table 4, Fig 2**).

Discussion

Clinical indications of liver dysfunction are regularly tested include liver enzyme plasma levels as AST and ALT. Higher risk of HCC and the existence of hepatocellular predominant diseases may be indicated by elevated ALT and AST values [16]. Li et al. found that HCC patients demonstrated higher levels of AST and ALT than chronic hepatitis patients [17] and this matched with current results revealed significant difference concerning AST (p < 0.0001) in HCC and LC compared to NDC groups but there was no significant difference between HCC and LC groups. Also, there was a significant difference between the three study groups in ALT suggesting that it might be an independent risk factor for the development of HCC in agrees with other reports [16,18].

APRI is a novel marker of liver cirrhosis and HCC patients' survival; also it provides additional prognostic insights in assessing the cirrhosis severity. APRI is associated with LC and HCC diagnosis among multiple high-risk populations [19]. APRI use to differentiate the cirrhotic from non-cirrhotic patients [20]. These studies support our findings as we found that there was a highly significant elevation of APRI in HCC and LC compared to NDC (p < 0.0001).

AAR is a validated diagnostic tool used to evaluate liver fibrosis [21, 22]. A significant correlation has been found between a high AAR and liver cirrhosis [23], particularly in individuals with non-alcoholic liver disease, where an AAR greater than 1.0 suggests the presence of cirrhosis. However, the AAR has not been shown to be a highly effective

predictor of HCC [17]. In current study, we found that median AAR was 1.385 in HCC group and this was agreed with Li et al, 2019 who found that AAR was 1.43 in HCC group [17]; also there was a significant difference between LC and NDC groups (p<0.0001), suggesting its potential role in disease severity of the liver.

A significant inflammatory burden often characterizes malignant diseases [24]. FIB-4 is a widely used as non-invasive scoring system helps in liver fibrosis assessment, a key risk factor in HCC as liver fibrosis and cirrhosis are associated with HCC. FIB-4 and liver cirrhosis have been correlated with the incidence of HCC and FIB-4 is known to be a good predictor of fibrosis and cirrhosis [25-29]. In current study, median FIB-4 in HCC group was 6.257 in agreed with Li et al. (2019) who found that FIB-4 was 6.66 in HCC group [17], also there was a highly significant difference (p<0.0001) in FIB-4 between LC and HCC compared to NDC but no significance difference was recorded between LC and HCC.

For inflammatory markers used in HCC diagnosis, we assessed PLR, CRP and NLR, which are commonly assessed through blood tests. These markers act to quantify the systemic inflammatory response linked to HCC and its potential impact on patient outcomes. All of these conditions are associated with inflammation, such as hepatocellular carcinoma [30]. The platelet-to-lymphocyte ratio (PLR) has been identified as a potential indicator of both malignancy and inflammatory conditions [31]. PLR is a well-established biomarker that assesses inflammatory and immune responses. It is a known prognostic factor in various malignancies, with elevated PLR values often correlating with unfavorable clinical outcomes in patients with HCC [32], While it is a useful marker, it's not typically used as a distinct diagnostic tool, however, we found that there was a significant difference between HCC and LC compared to NDC (P<0.05) but no significant difference between HCC and LC groups, in contrast to Chen et al. (2024) [33] who conducted that there was a significant difference in PLR between HCC patients and LC cirrhosis (p < 0.0001).

CRP is acute-phase reactant produced by liver cells in response to inflammation [34]. Blood CRP levels are increased due to inflammation occurred with the incidence of various cancers, including HCC [35]. Even though it found in very high values in cirrhotic patients and secreted in the presence of HCC, it isn't a diagnostic marker for HCC [36,37]. however it is still been noted to have significant prognostic value [38-40]. Currently, CRP showed highly significant difference in HCC and LC compared to NDC (p<0.05).

The neutrophil to lymphocyte (NLR) ratio in the peripheral blood [41] has also been proven to be a valuable indicator of the inflammatory state and a predictor of clinical survival in HCC [42, 43] and it has already been proven to be associated with a poor prognosis across a range of tumors [44]. Currently, we found a highly significant difference in NLR in HCC and LC groups compared to NCD group. However its ability to distinguish HCC from LC is limited. This means that NLR isn't specific enough in HCC diagnosis.

Alfa-fetoprotein (AFP) is a fetal-specific glycoprotein produced by the fetus's liver and its synthesis is suppressed in adult [10]. Currently, serum AFP expression levels were significantly higher in HCC group compared to LC and NDC group in agree with other reports [45,46].

Gamma-glutamyltransferase (GGT) is a cell membrane-bound enzyme secreted in healthy adults by endothelial cell of bile duct and hepatic Kupffer cell and its activity increases in fetal liver and HCC [47]. Several studies reported that serum GGT was effective diagnostic biomarker of hepatobiliary disease and various tumors [48,49]. GGT plays a role in the growth and development of HCC [50,51]. GGT levels abnormally increase in other liver diseases such as a viral hepatitis, alcoholic

hepatitis, and liver cirrhosis, so it can't be used as effective indicator for HCC screening [42]. In current study, GGT showed higher activity in LC and HCC compared to NDC group (p<0.05), although higher levels of GGT in HCC compared to LC was detected non significant. GGT was associated with ALT and Also. AFP Pt count. was significantly correlated with AST, ALT, Pt and NLR. Regression analysis showed AFP, ALT, Pt and INR displayed substantial prediction for tumor development. The diagnostic performance of AFP, ALT, and Pt using ROC curve analysis demonstrated only AFP with excellent accuracy (90.2%) and AUC of 0.972 supporting its clinical applicability.

Conclusion

Non-invasive inflammatory markers an fibrotic indices are involved in disease severity of the liver. ALT, Pt and INR displayed substantial prediction for tumor development and could be used as early predictors for HCC.

Table (2) Correlation between AFP, GGT and AST with other biomarkers in all patients groups

AFP							
Variable	Rho	P value					
AST	0.253	0.022					
ALT	0.350	0.001					
NLR	-0.285	0.009					
Pt	0.255	0.021					
	GGT						
ALT	0.311	0.004					
Pt	0.229	0.038					
	AST						
ALT	0.682	< 0.0001					
AAR	-0.599	< 0.0001					
APRI	0.680	< 0.0001					
FIB4	0.457	< 0.0001					
NLR	-0.350	0.001					
PLR	-0.306	0.005					
AFP	0.253	0.022					

Data was presented by Spearman correlation Coefficient test

Table (3) Regression Analysis of AFP, GGT, ALT, Pt and INR as independent predictors in HCC development

Predictor	В	SE	P value	95% CI	OR
AFP	-0.209	0.048	< 0.0001	0.739-0.891	0.812
GGT	-0.005	0.006	0.392	0.983-1.007	0.995
ALT	-0.032	0.015	0.03	0.940-0.097	0.968
Pt	-0.018	0.007	0.014	0.969-0.996	0.982
INR	1.546	0.812	0.057	0.956-23.027	4.693

Table (4) Diagnostic performance of AFP, for discriminating HCC from LC

Variable	AUC	P value	Cut-off	95% CI	Sens.	Spec.	LR+	LR-	PPV	NPV	Accuracy
AFP	0.972	≤0.0001	18.5	0.00-1.0	81.1	95.6	18.43	0.197	93.9	0.877	90.2
ALT	0.673	0.007	21.5	0.557-0.789	83.8	40.0	1.39	0.405	53.4	75.0	59.8
Pt	0.634	0.038	46.0	0.511-0.756	75.7	40.0	1.26	0.607	50.9	66.7	56.1

AUC; Area under ROC curve, PPV; Positive predictive value, NPV; Negative predictive value

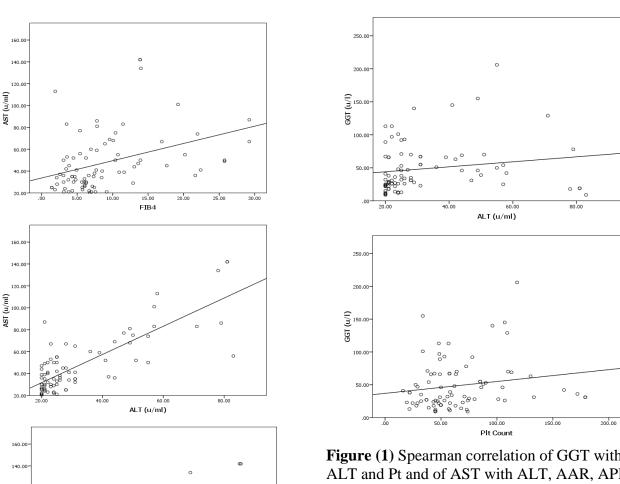
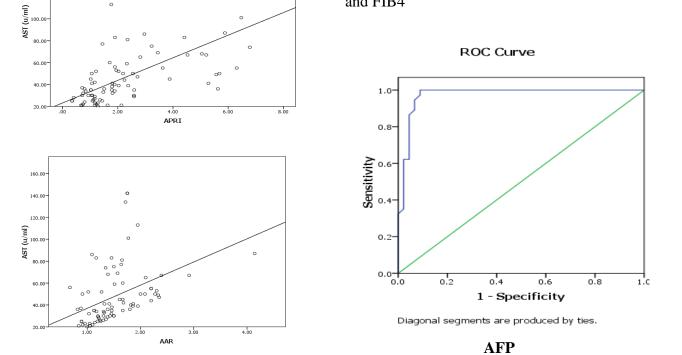
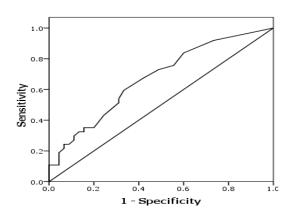


Figure (1) Spearman correlation of GGT with ALT and Pt and of AST with ALT, AAR, APRI and FIB4







Diagonal segments are produced by ties.

ALT

Figure (2) ROC curve of AFP, ALT

4. References

- 1. Zhang J, He X, Wan Y, et al. (2021). CD44 promotes hepatocellular carcinoma progression via upregulation of YAP. ExpHematolOncol.;10(1):54.
- 2. Mima K, Okabe H, Ishimoto T, Hayashi H, Nakagawa S, Kuroki H, Watanabe M, Beppu T, Tamada M, Nagano O, Saya H, Baba H (2012). CD44s regulates the TGF-beta-mediated mesenchymal phenotype and is associated with poor prognosis in patients with hepatocellular carcinoma. Cancer Res 72: 3414–3423.
- 3. Ferlay J, Ervik M, Lam F, Laversanne M, et al. (2024). Global Cancer Observatory: Cancer Today. Lyon, France: International Agency for Research on Cancer.
- 4. Liu M, Jiang L, Guan XY. (2014) The genetic and epigenetic alterations in human hepatocellular carcinoma: a recent update. Protein Cell.:5:673–691.
- 5. Gomaa AI, Khan SA, Toledano MB, Waked I, Taylor-Robinson SD. (2008) Hepatocellular carcinoma: epidemiology, risk factors and pathogenesis. *World J Gastroenterol.*;**14**(27):4300-4308. doi:10.3748/wjg.14.4300
- 6. Poon D, Anderson BO, Chen LT, et al. Management of hepatocellular carcinoma in Asia: consensus statement from the Asian Oncology Summit (2009). Lancet Oncol.:10(11):1111-1118.
- 7. Pascut, D., Pratama, M.Y., &Tiribelli, C. (2020) HCC occurrence after DAA treatments: molecular tools to assess the

- posttreatment risk and surveillance., Taylor & Francis. p. HEP21.
- 8. Zhao, Y.-J., Ju, Q., & Li, G.-C. (2013) Tumor markers for hepatocellular carcinoma. Molecular and clinical oncology.; **1(4)**: 593-598.
- 9. Ho WH, Lee KT, Chen HY, Ho TW, Chiu HC. (2012) Disease-free survival after hepatic resection in hepatocellular carcinoma patients: a prediction approach using artificial neural network. PLoS One.;7(1):e29179.

 doi:10.1371/journal.pone.0029179
- 10. Zhou L, Liu J, Luo F. (2006) Serum tumor markers for detection of hepatocellular carcinoma. *WorldJGastroenterol.*;**12(8)**:1 175-1181. doi:10.3748/wjg.v12.i8.1175
- 11. Cancer Clinical Trial Eligibility Criteria: Patients with HIV, Hepatitis B Virus, or Hepatitis C Virus Infections Guidance for Industry. (2020)
- 12. Wai CT, Greenson JK, Fontana RJ, Kalbfl eisch JD, Marrero JA, Conjeevaram HS, L ok AS (2003) A simple non-invasive index can predict both significant fibrosis and cirrhosis in patients with chronic hepatitis C. Hepatology; 38:518–26.
- 13. .Vallet-Pichard A, Mallet V, Nalpas B, Verkarre V, Nalpas A, Dhalluin-Venier V, Fontaine H, Pol S . (2007) FIB-4: an inexpensive and accurate marker of fibrosis in HCV infection. Comparison with liver biopsy and FibroTest. Hepatology;**46**:32–36.
- 14. Proctor MJ, Morrison DS, Talwar D, Bal mer SM, Fletcher CD, O'Reilly DS, Fouli s AK, Horgan PG, McMillan DC . (2011) A comparison of inflammation-based prognostic scores in patients with cancer. A Glasgow Inflammation Outcome Study. Eur J Cancer;47:2633–2641
- 15. Nahm FS. (2022) Receiver operating characteristic curve: overview and practical use for clinicians. *Korean J Anesthesiol.*; **75(1)**:25-36.
- 16. Qin S, Wang J, Yuan H, He J, Luan S, Deng Y. (2024) Liver function indicators and risk of hepatocellular carcinoma: a bidirectional mendelian randomization study. Front Genet.;14:1260352.
- 17. Li X, Xu H, Gao P. (2019) Fibrosis Index Based on 4 Factors (FIB-4) Predicts Liver

- Cirrhosis and Hepatocellular Carcinoma in Chronic Hepatitis C Virus (HCV) Patients. Med SciMonit.;25:7243-7250.
- 18. Ogasawara N, Saitoh S, Akuta N, et al. (2020) Advantage of liver stiffness measurement before and after direct-acting antiviral therapy to predict hepatocellular carcinoma and exacerbation of esophageal varices in chronic hepatitis C. Hepatol Res;50(4):426-438.
- 19. Allenson K, Roife D, Kao LS, Ko TC, Wray CJ. (2020) Estimation of hepatocellular carcinoma mortality using aspartate aminotransferase to platelet ratio index. *JGastrointestOncol*.;**11**(2):291-297.
- 20. Abdelgawad IA. (2015) Clinical utility of simple non-invasive liver fibrosis indices for predicting hepatocellular carcinoma (HCC) among Egyptian patients. *J ClinPathol.*;68:154-60.
- 21. Sheth SG, Flamm SL, Gordon FD, Chopra S. (1998) AST/ALT ratio predicts cirrhosis in patients with chronic hepatitis C virus infection. *Am J Gastroenterol.*;**93**:44–48.
- 22. Giannini E, Botta F, Fasoli A, et al. (1999) Progressive liver functional impairment is associated with an increase in AST/ALT ratio. Dig Dis Sci.;44:1249–53.
- 23. Williams AL, Hoofnagle JH: (1988) Ratio of serum aspartate to alanine aminotransferase in chronic hepatitis. Relationship to cirrhosis. Gastroenterology; **95**: 734–39.
- 24. Sit M, Aktas G, Ozer B, et al. (2019) Mean platelet volume: an overlooked herald of malignant thyroid nodules. ActaClinicaCroatica.;58(3):417-420.
- 25. Yu ML, Lin SM, Lee CM et al. (2006) A simple noninvasive index for predicting long-term outcome of chronic hepatitis C after interferon-based therapy. Hepatology (Baltimore, Md); 44: 1086–97
- 26. Masuzaki R, Tateishi R, Yoshida H et al. (2009) Prospective risk assessment for hepatocellular carcinoma development in patients with chronic hepatitis C by transient elastography. Hepatology (Baltimore, Md); **49**: 1954–61.
- 27. Nunes D, Fleming C, Offner G et al. (2010) Noninvasive markers of liver

- fibrosis are highly predictive of liver-related death in a cohort of HCV-infected in dividuals with and without HIV infection. *Am J Gastroenterol*; **105**: 1346–53
- 28. Vergniol J, Foucher J, Terrebonne E et al. (2011) Noninvasive tests for fibrosis and liver stiffness predict 5-year outcomes of patients with chronic hepatitis C. Gastroenterology; **140**: 1970–79, 9.e1-3.
- 29. Park LS, Tate JP, Justice AC et al. (2011) FIB-4 index is associated with hepatocellular carcinoma risk in HIV-infected patients. Cancer Epidemiol Biomarkers Prev; **20**: 2512–17.
- 30. Sanghera C, Teh JJ. (2019) The systemic inflammatory response as a source of biomarkers and therapeutic targets in hepatocellular carcinoma.Liver International.;39(August):1-16.
- 31. Meryem B, Tel A. (2021) Department of Internal
 Medicine.BoluAbantIzzetBaysalUniversit
 y, Medical School, Bolu, Turkey
 Department of Gastroenterology,
 BoluAbantIzzetBaysal University,
 Medical School, Bolu, Turkey.;4(2):14853
- 32. He C, Zhang Y, Cai Z, Lin X. (2019) The prognostic and predictive value of the combination ofthe neutrophil-tolymphocyte ratio and the platelet-tolymphocyte ratio in patients with hepatocellular carcinoma who receive transarterial chemoembolization therapy. Cancer Manag Res.: 11(2):1391-1400.
- 33. Chen X, Mohammed AF, Li C. (2024)
 Assessment of the Clinical Value of
 Platelet-to-Lymphocyte Ratio in Patients
 with Hepatocellular Carcinoma.
 ClinApplThrombHemost.;30:1076029623
 1221535.
- 34. Castell JV, Gomez-Lechon MJ, David M, Fabra R, Trullenque R, Heinrich PC. (1990) Acute-phase response of human hepatocytes: regulation of acute-phase protein synthesis by interleukin-6. Hepatology.; **12**: 1179–1186.
- Carr BI, Akkiz H, Guerra V, Üsküdar O, Kuran S, Karaoğullarından Ü, et al. (2018) C-reactive protein and hepatocellular carcinoma: analysis of its

- relationships to tumor factors. ClinPract (Lond).;**15**(Spec Issue):625-634.
- 36. Fabris C, Pirisi M, Soardo G, et al. (1996) Diagnostic usefulness of acute-phase protein measurement in hepatocellular carcinoma. Cancer Invest.;**14**:103–108.
- 37. Lin ZY, Wang LY, Yu ML, et al (2000). Role of serum C-reactive protein as a marker of hepatocellular carcinoma in patients with cirrhosis. *J GastroenterolHepatol.*;**15(4)**:417-421. doi:10.1046/j.1440-1746.2000.02149.x
- 38. Hashimoto K, Ikeda Y, Korenaga D, Tanoue K, Hamatake M. (2005) The impact of preoperative serum C-reactive protein on the prognosis of patients with hepatocellular carcinoma. Cancer.; **103**:1856–1864. doi: 10.1002/cncr.20976.
- 39. Shin JH, Kim CJ, Jeon EJ, et al. (2015) Overexpression of C-reactive Protein as a Poor Prognostic Marker of Resectable Hepatocellular Carcinomas. *J* PatholTransl Med.:49:105–111.
- 40. Sieghart W, Pinter M, Hucke F, et al. (2013) Single determination of C-reactive protein at the time of diagnosis predicts long-term outcome of patients with hepatocellular carcinoma. Hepatol.;57:2224–2234.
- 41. Buonacera A, Stancanelli B, Colaci M, Malatino L. (2022) Neutrophil to Lymphocyte Ratio: An Emerging Marker of the Relationships between the Immune System and Diseases. *Int J Mol Sci.*;23(7):3636
- 42. Zheng J, Cai J, Li H, et al. (2017) Neutrophil to Lymphocyte Ratio and Platelet Lymphocyte to Ratio Prognostic Predictors for Hepatocellular Carcinoma **Patients** with Various Treatments: a Meta **Analysis** and **Systematic** Review. Cell PhysiolBiochem.; 44:967–981.
- 43. Yu Y, Song J, Zhang R, et al. (2017) Preoperative neutrophil-to-lymphocyte ratio and tumor-related factors to predict microvascular invasion in patients with hepatocellular carcinoma. Oncotarget.2017; 8:79722–79730.
- 44. Templeton AJ, McNamara MG, Šeruga B, Vera-Badillo FE, Aneja P, Ocaña A, et al.

- (2014) Prognostic role of neutrophil-to-lymphocyte ratio in solid tumors: a systematic review and meta-analysis. *J Natl Cancer Inst.*; **106**: dju124.
- 45. Abdel-Hafiz SM, Hamdy HE, Khorshed FM, et al. (2018) Evaluation of Osteopontin as a Biomarker in Hepatocellular Carcinomas in Egyptian Patients with Chronic HCV Cirrhosis. Asian Pac *J Cancer Prev.*;**19**(**4**):1021-1027. doi: 10.22034/APJCP.2018.19.4.1021.
- 46. Mostafa EF, Eltaher HM, Nasr Eldin E, Hassany SM.(2023) Diagnostic Accuracy of Plasma Osteopontin in Egyptian Hepatocellular Carcinoma Patients. Afro-Egyptian *Journal of Infectious and Endemic Diseases.*; **13(1)**, 3-14.
- 47. Cui R, He J, Zhang F, Wang B, Ding H, Shen H, Li Y, Chen X. (2003) Diagnostic value of protein induced by vitamin K absence (PIVKAII) and hepatoma-specific band of serum gamma glutamyltransferase (GGTII) as hepatocellular carcinoma markers complementary to alphafetoprotein. *Br J Cancer.*; **88**: 1878-1882
- 48. Zhao WC, Fan LF, Yang N, et al. (2013) Preoperative predictors of microvascular invasion in multinodular hepatocellular carcinoma. *Eur J SurgOncol*;**39**:858–64.
- 49. Dalpiaz O, Pichler M, Mrsic E, et al. (2015) Preoperative serum-gamma glutamyltransferase (GGT) doesnot represent anindependentprognostic factor in a European cohort of patients with nonmetastatic renal cell carcinoma. *J ClinPathol.*;68:547–51.
- 50. Corti A, Franzini M, Paolicci A, Pompella A (2010) Gamma-glutamyltransferase of cancer cells at the crossroads of tumor progression, drug resistance and drug targeting. Anticancer Res **30**: 1169–1182.
- 51. Hanigan MH (2014) Gamma-GlutamylTranspeptidase: Redox Regulation and Drug Resistance. Adv Cancer Res 122: 103–141.
- 52. Faber W, Sharafi S, Stockmann M, et al. (2013) Long-term results of liver resection for hepatocellular carcinoma in noncirrhotic liver. Surgery.;153:510e517.